

Fiber orientations from diffusion MRI and histology in the macaque brain

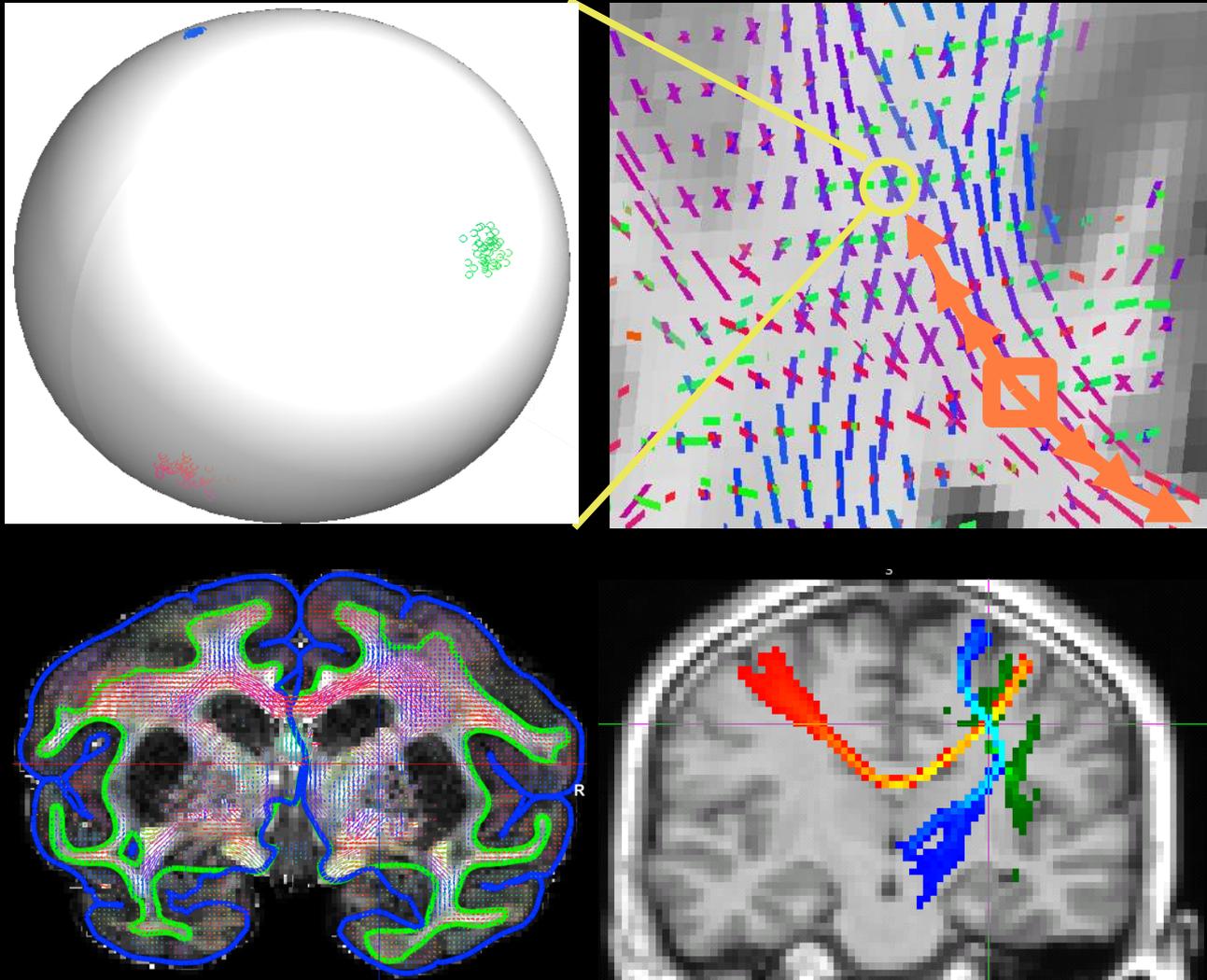
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Dikranian¹, David C. Van Essen¹, and Matthew F. Glasser¹



2

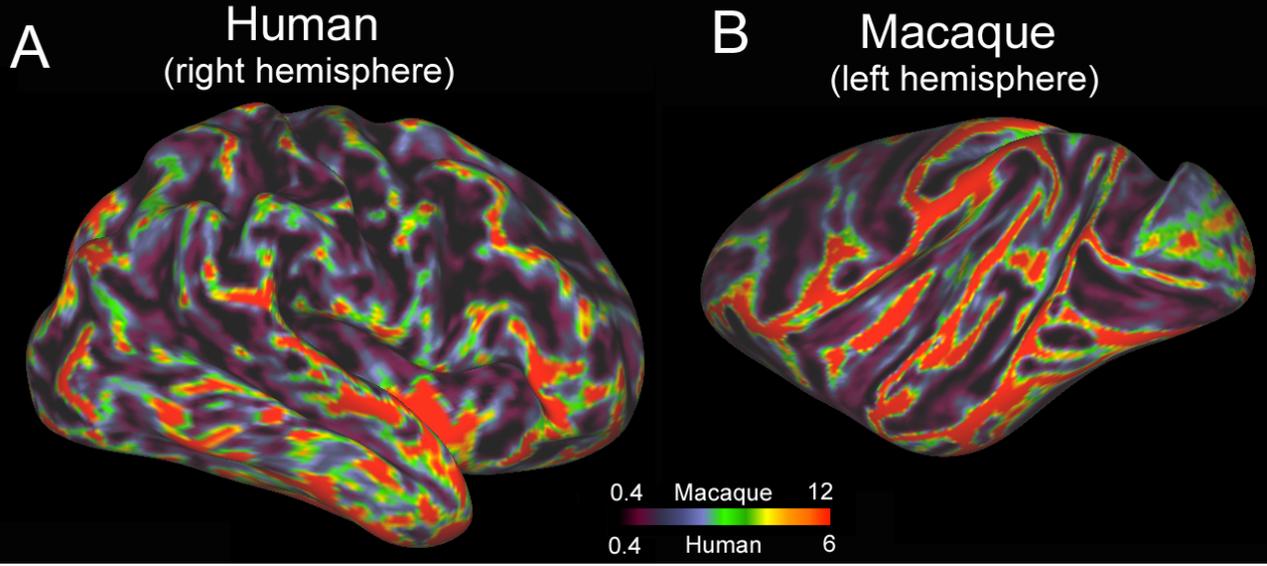


Diffusion MRI and tractography

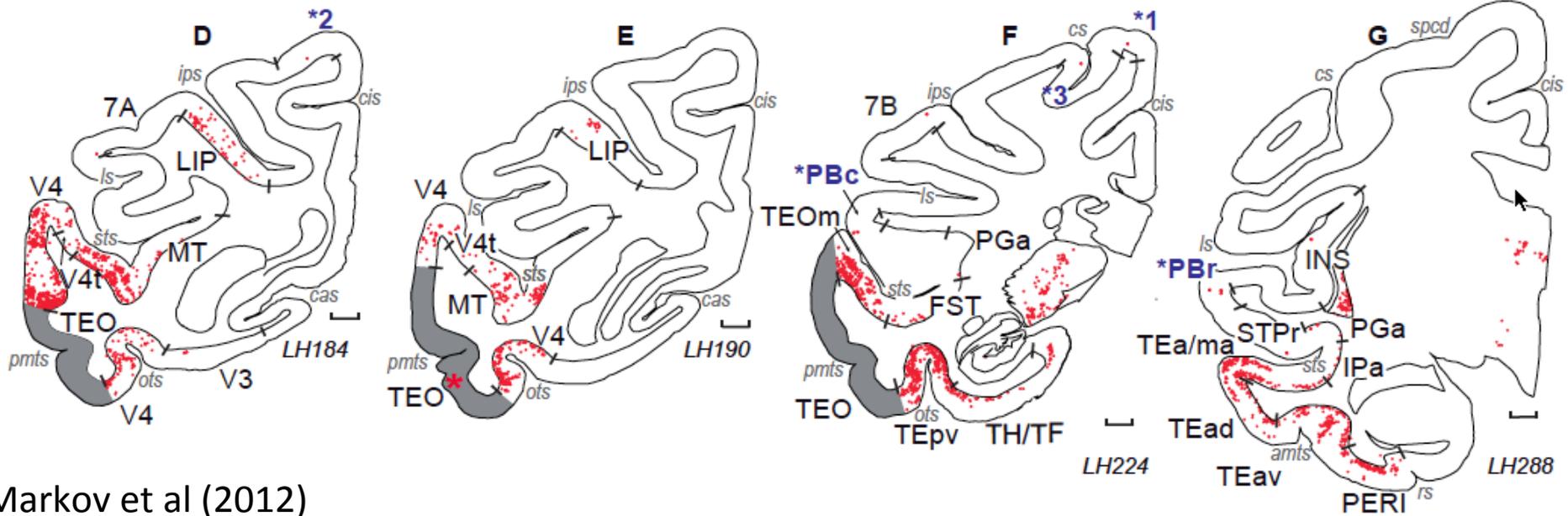


Gyral bias in tractography streamline “density”

- Top : Streamline density maps from human and post-mortem macaque
- Bottom : Tracer injections from macaque

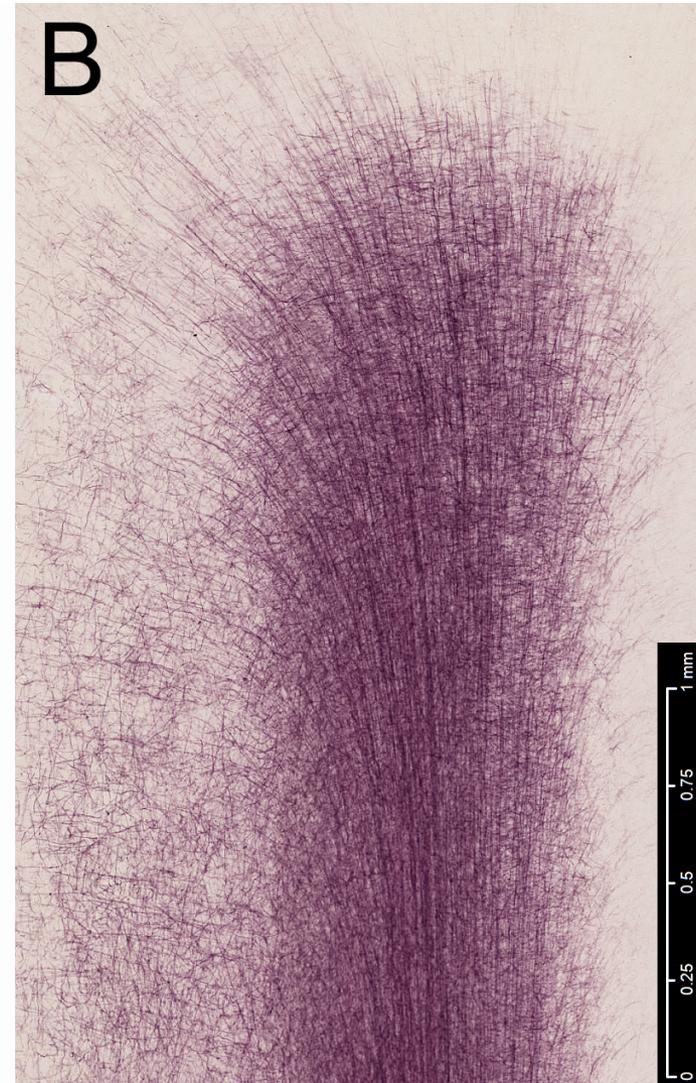
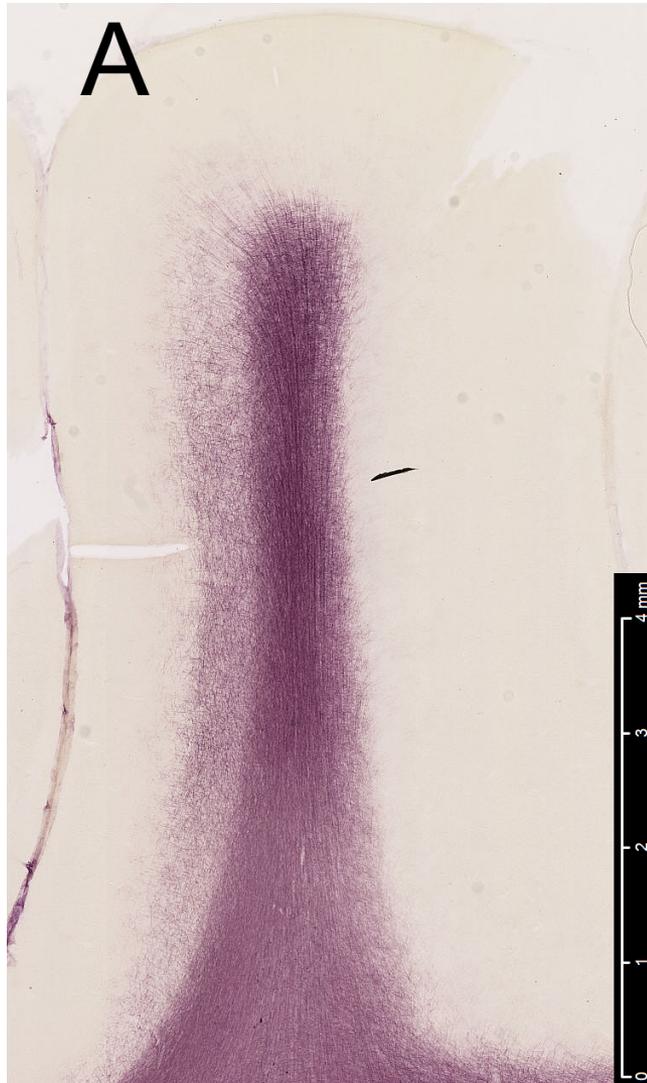


Van Essen et al (2013), in press

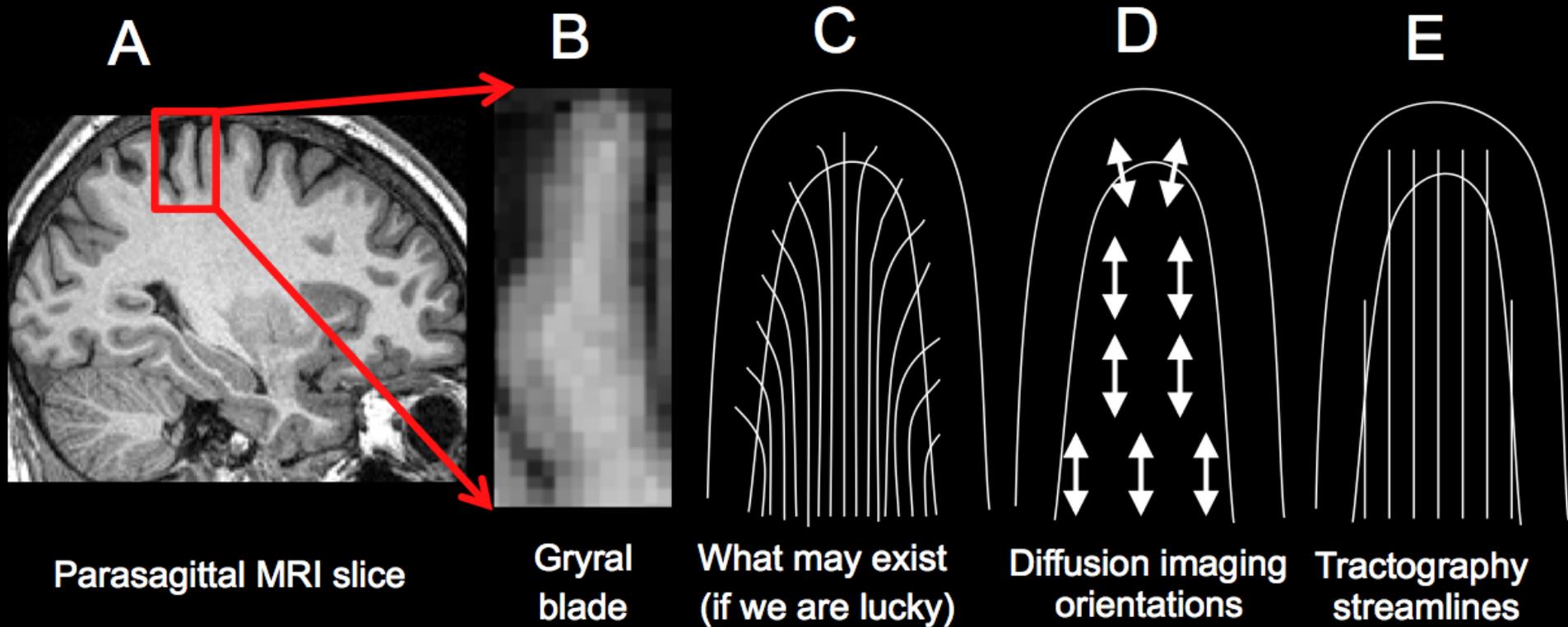


Markov et al (2012)

Insights from histology

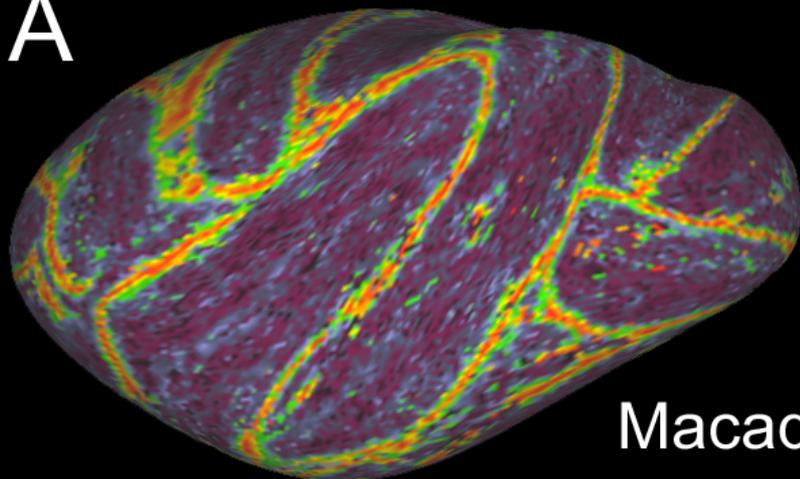


Tractography predictions

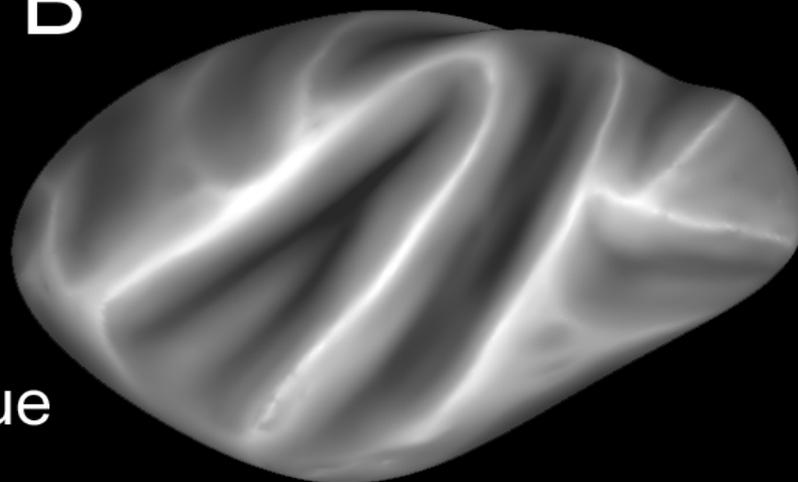


Fiber orientation closest
to surface normal

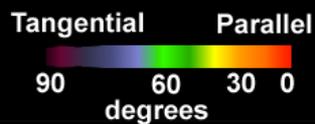
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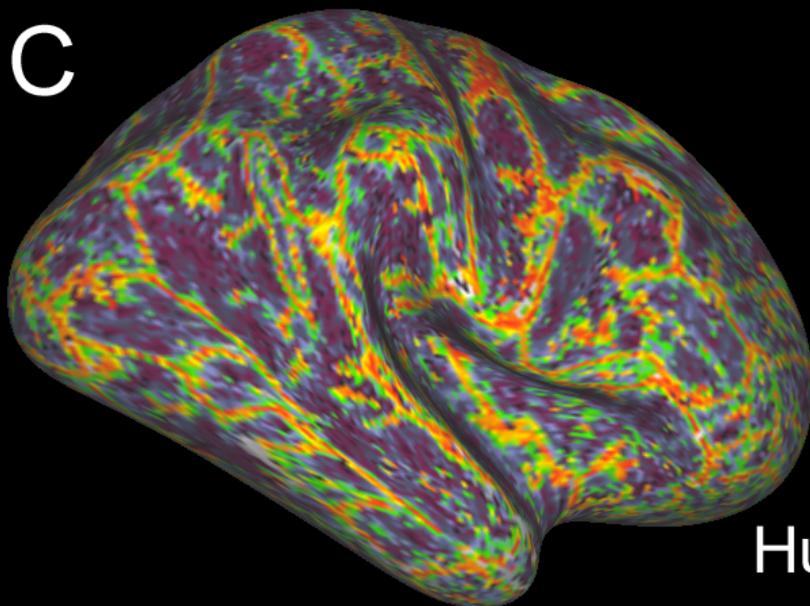
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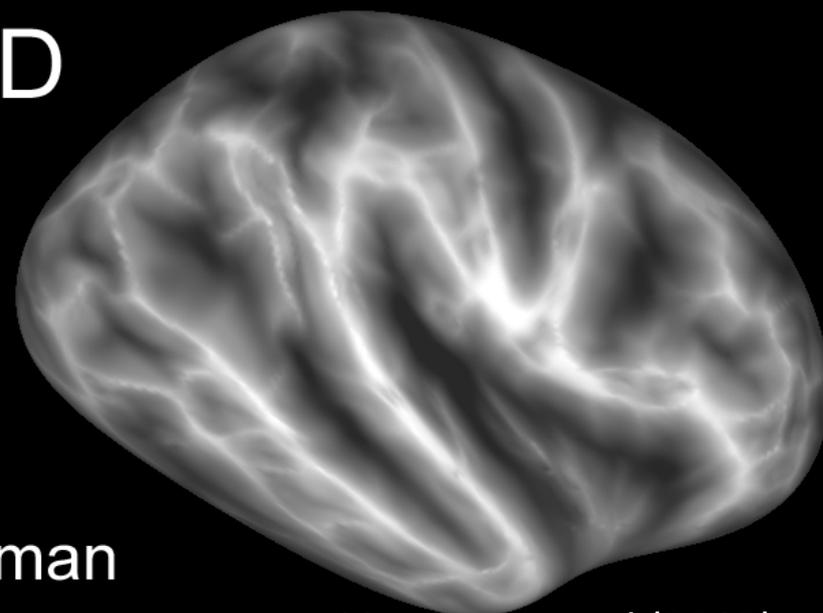
Macaque



C

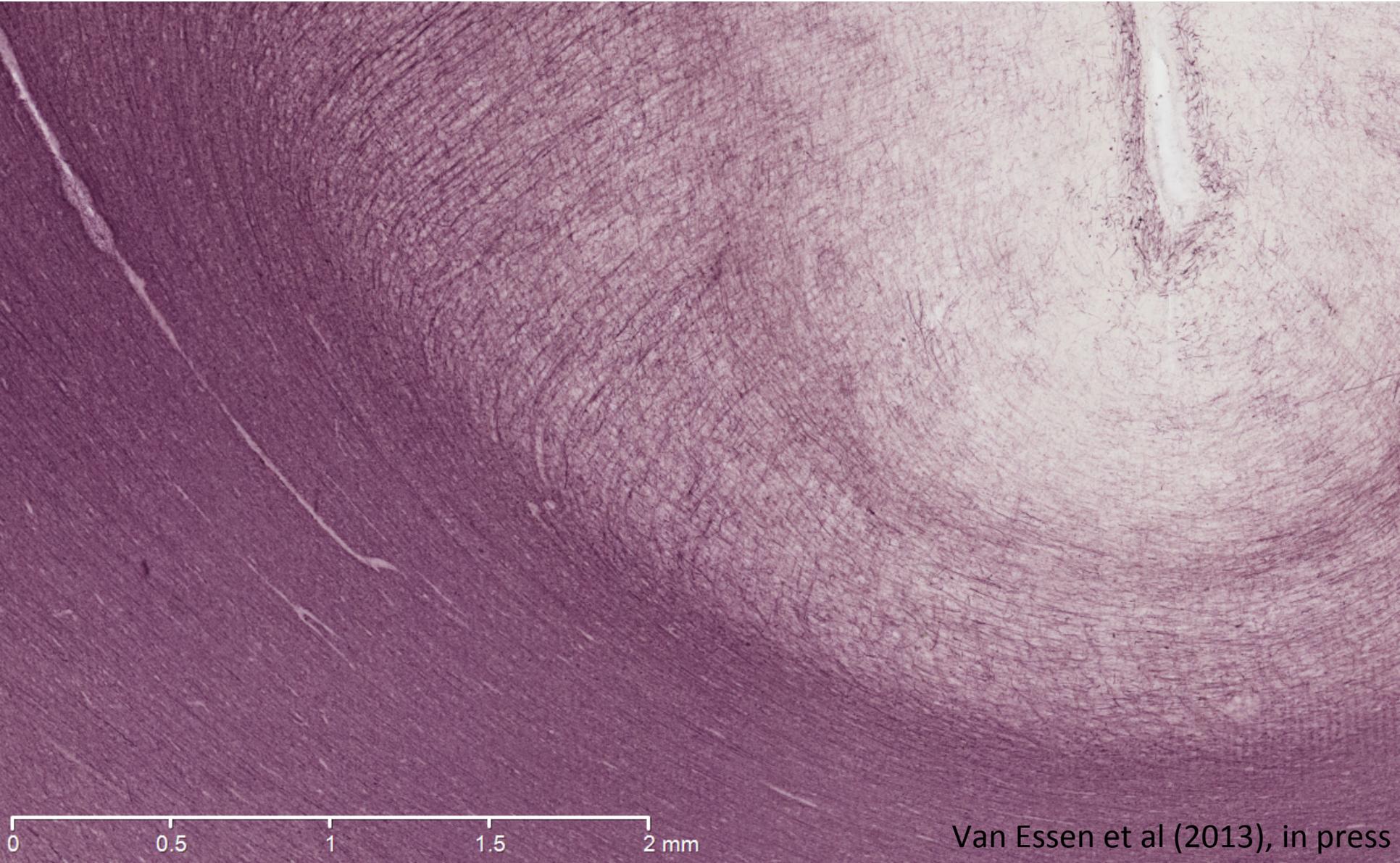


D

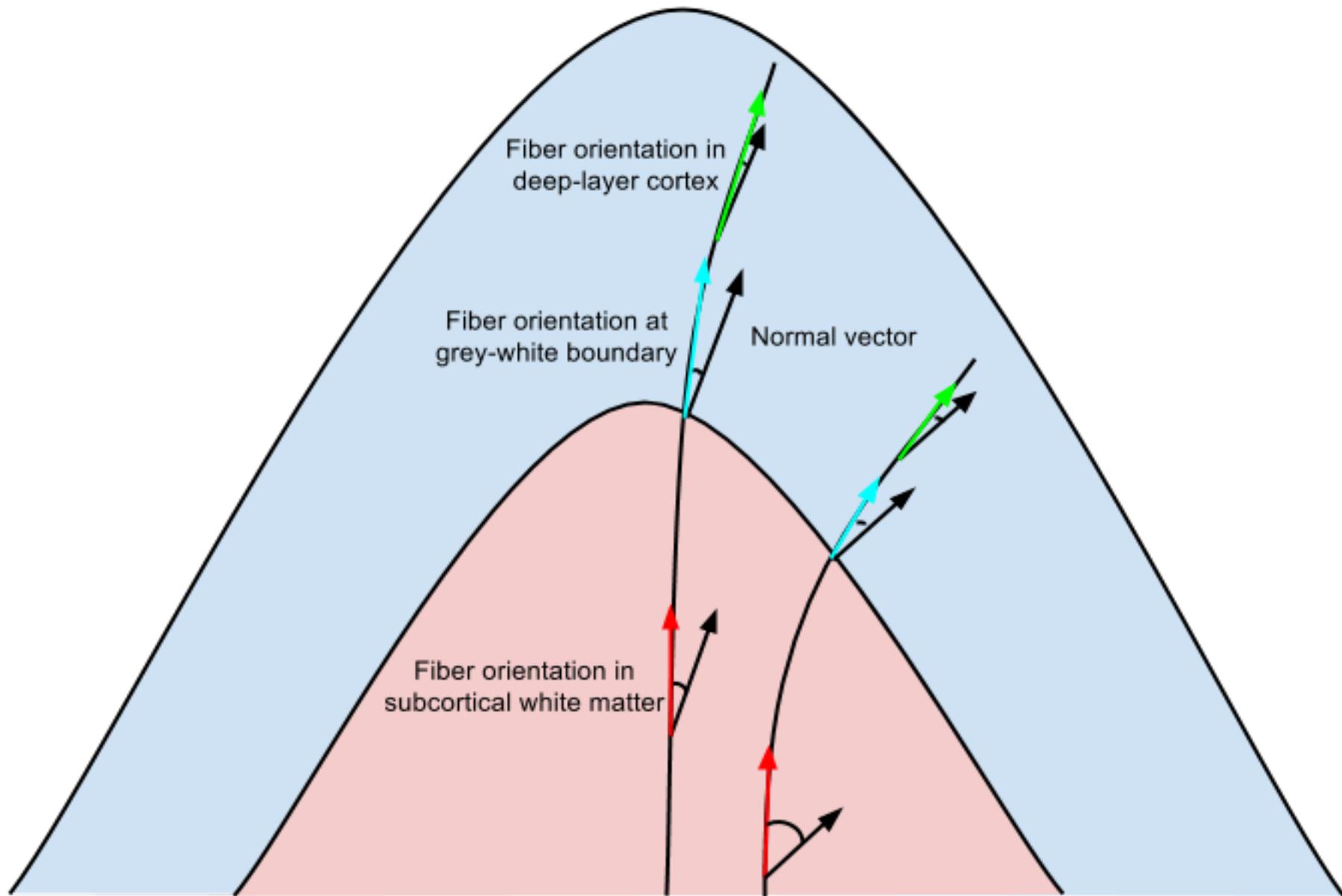


Human

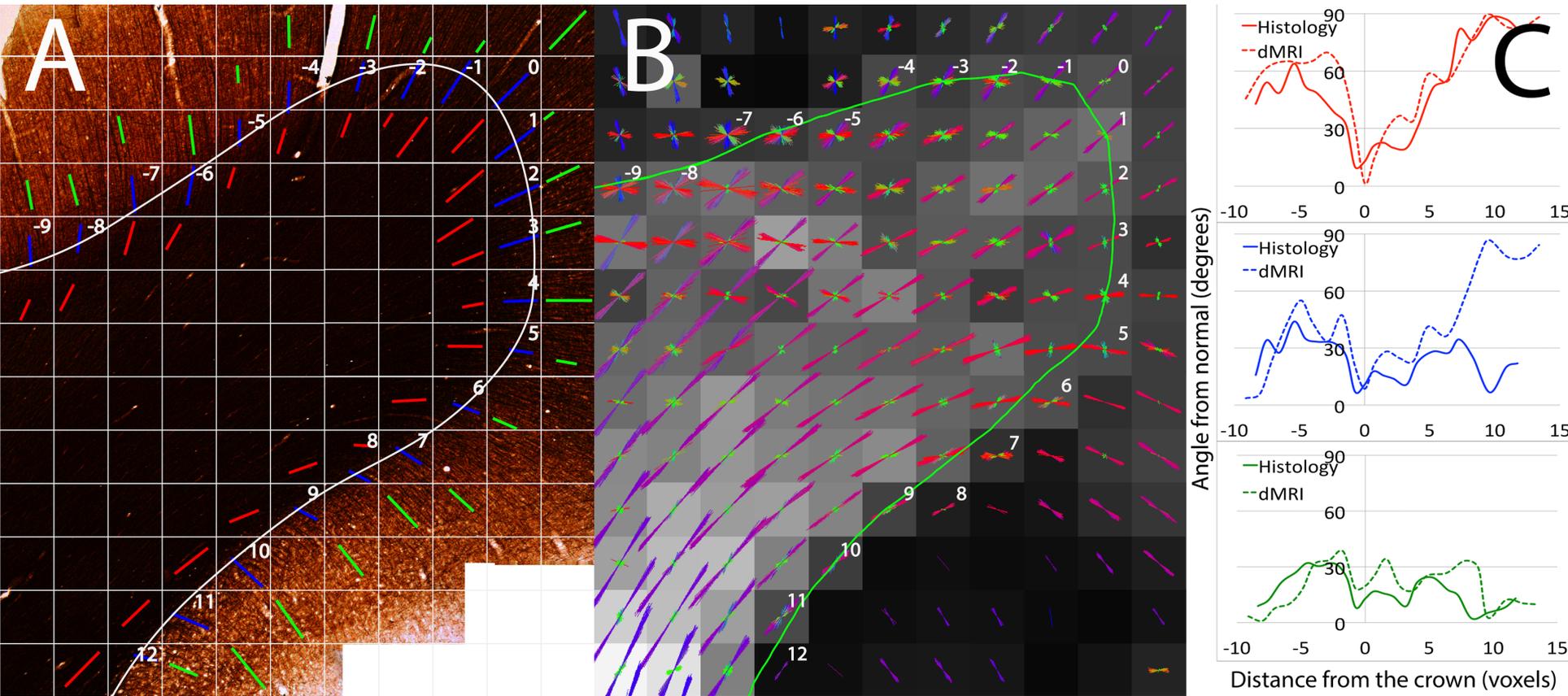
Insights from histology



Fiber orientations near cortex



Comparing DTI and histology



Summary

- Diffusion MRI and tractography are powerful tools for generating connectomes
- However, they suffer from technical limitations, such as resolving gyral biases
- Tractography algorithms can be informed through histological data
- Fiber estimates can only be improved through better acquisition

Supplementary Slides

Imaging Methods

➤ Histology

A **postnatal day 6 macaque brain**. Sections were immunostained with antibody to **myelin basic protein (MBP, MAB395, Millipore)** and scanned on a NanoZoomer 2 (Hamamatsu) scanning microscope equipped with Olympus lens at 20X (**0.9225 $\mu\text{m} \times 0.9225 \mu\text{m}^2$ resolution**).

A modified Gallyas myelin stained section from an **adult macaque brain** was also digitized in a similar fashion*.

➤ Post-mortem Diffusion MRI**

A diffusion-weighted MRI dataset of a perfusion-fixed **adult macaque brain** was acquired using a 4.7 T Bruker scanner.

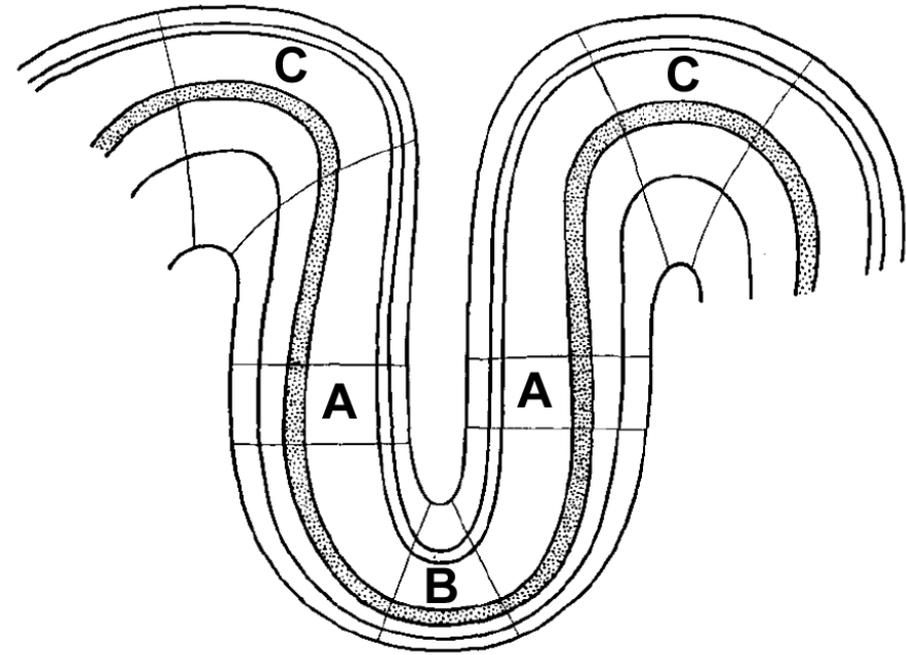
Scans were performed using a 3D multi-shot, spin-echo sequence (with in-plane **resolution $430 \times 430 \mu\text{m}^2$** , TE = 33 ms, TR = 350 ms)

120 DW directions at $b=8000 \text{ s/mm}^2$, 17 $b=0 \text{ s/mm}^2$, 128 slices with a thickness of 430 μm .

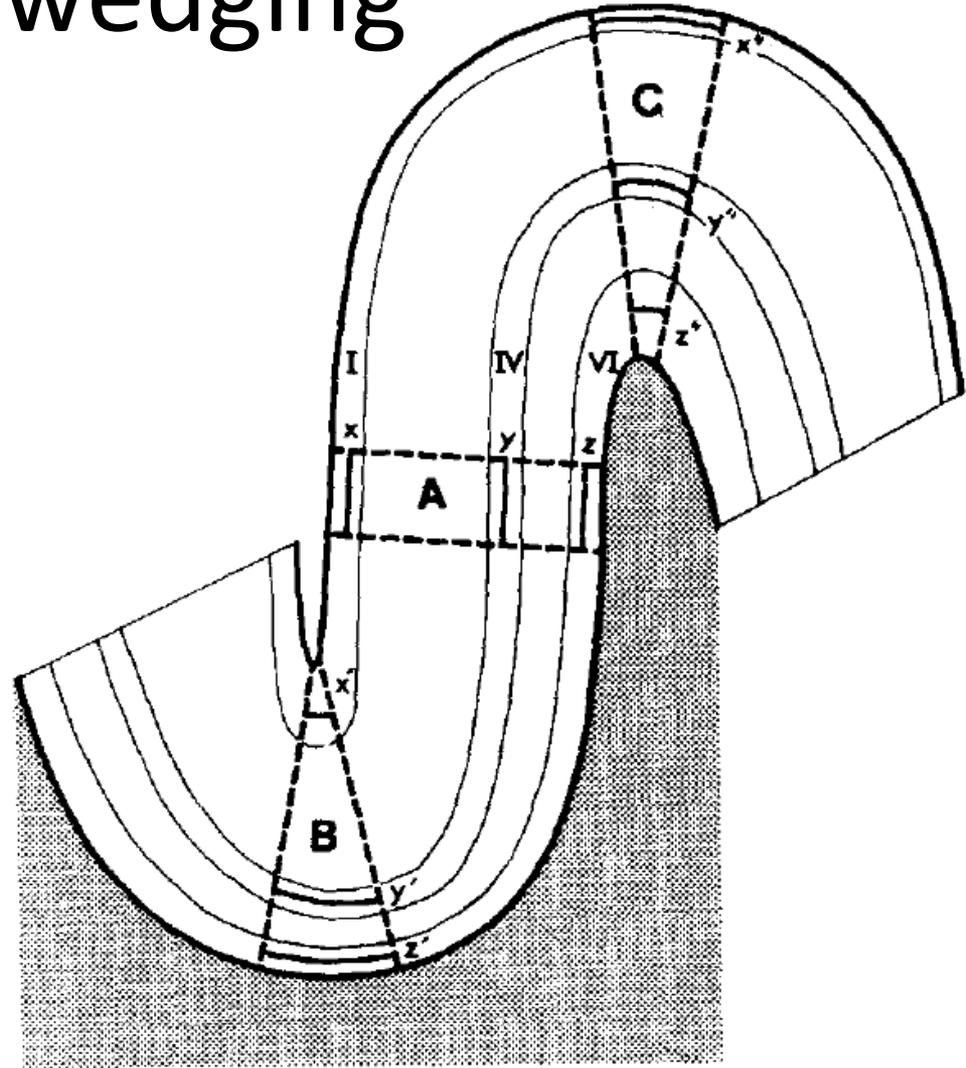
* Data are courtesy of JL Price, WashU, School of Medicine

** Data from [D'Arceuil et al, NeuroImage 35:553-565, 2007]

Cortical wedging

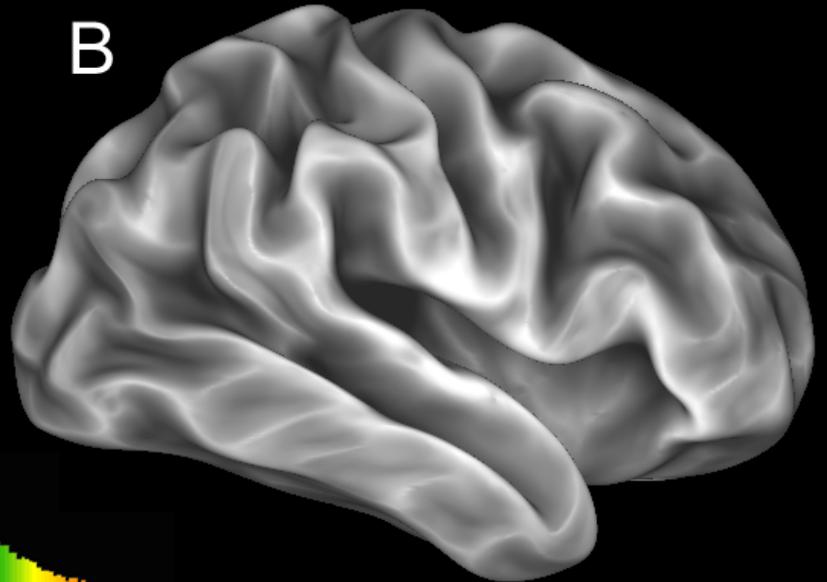
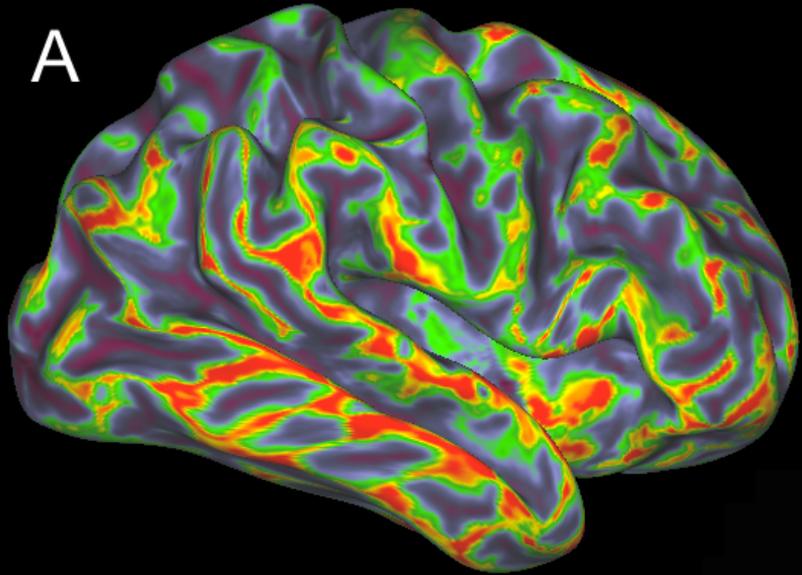


Bok (1929) (via Waehnert et al., 2013)

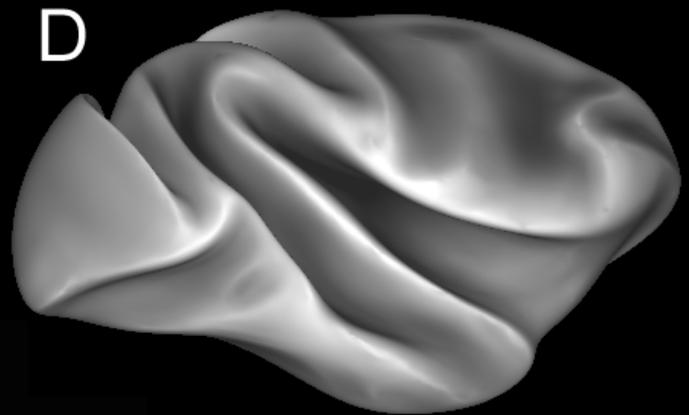
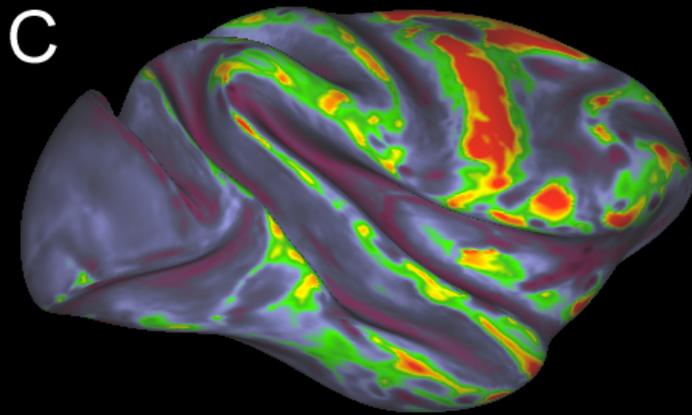
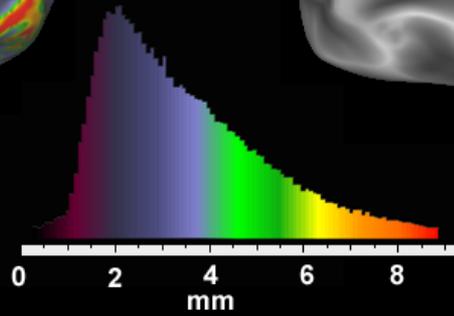


Van Essen and Maunsell, (1980)

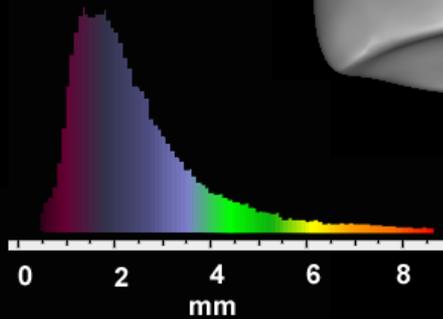
Gyral vs sulcal wedges: Cortical volume per unit area of gray/white surface



Human

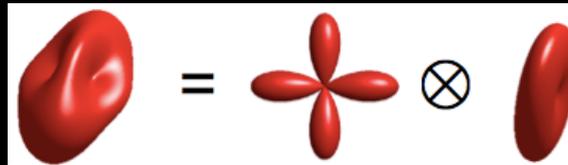


Macaque



- Assuming mono-exponential decay in q-space:
[Behrens et al, MRM 2003], [Kaden et al, NeuroImage 2007]

$$S_k = S_0 \left[(1 - f) \exp(-b_k d) + f \int_0^{2\pi} \int_0^{\pi} H(\theta, \phi) \exp(-b_k d (\mathbf{g}_k^T \mathbf{v})^2) \sin \theta d\theta d\phi \right]$$



- If the fODF is modelled as a Delta function (or sum of Delta functions), we get the **ball & stick model** [Behrens et al, MRM 2003, NeuroImage 2007]:

$$S_k = S_0 \left[(1 - f) \exp(-b_k d) + f \exp(-b_k \mathbf{g}_k^T \mathbf{v})^2 \right]$$

Structure Tensor Analysis

Given an image $I(x, y)$ and its spatial gradient vector

$$\nabla I = [I_x \quad I_y]^T$$

← spatial partial derivative along y
(Gaussian smoothed)

The 2x2 *gradient tensor* is: $Q = \nabla I \cdot \nabla I^T = [q_{ij}]$

The 2x2 *structure tensor* is: $S = [s_{ij}], \quad s_{ij} = g_{\sigma,w} * \{q_{ij}\}$

← Gaussian filter with window size w
and spatial scale σ

The eigenvector of the structure tensor associated with the smallest eigenvalue gives the *coherence* direction.

Comparing DTI and Histology

